

Scotland's Rural College

## **Contribution of the drought tolerance-related stress-responsive NAC1 transcription factor to resistance of barley to Ramularia leaf spot**

McGrann, GRD; Steed, A; Burt, C; Goddard, R; Lachaux, C; Bansal, A; Corbitt, M; Gorniak, K; Nicholson, P; Brown, JKM

*Published in:*  
Molecular Plant Pathology

*DOI:*  
[10.1111/mpp.12173](https://doi.org/10.1111/mpp.12173)

First published: 01/01/2014

*Document Version*  
Publisher's PDF, also known as Version of record

[Link to publication](#)

### *Citation for pulished version (APA):*

McGrann, GRD., Steed, A., Burt, C., Goddard, R., Lachaux, C., Bansal, A., Corbitt, M., Gorniak, K., Nicholson, P., & Brown, JKM. (2014). Contribution of the drought tolerance-related stress-responsive NAC1 transcription factor to resistance of barley to Ramularia leaf spot. *Molecular Plant Pathology*, 16(2), 201 - 209. <https://doi.org/10.1111/mpp.12173>

### **General rights**

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

### **Take down policy**

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

# Contribution of the drought tolerance-related *Stress-responsive NAC1* transcription factor to resistance of barley to *Ramularia* leaf spot

GRAHAM R. D. MCGRANN<sup>1,\*†</sup>, ANDREW STEED<sup>1</sup>, CHRISTOPHER BURT<sup>1,‡</sup>, RACHEL GODDARD<sup>1</sup>, CLEA LACHAUX<sup>1</sup>, ANURADHA BANSAL<sup>1</sup>, MARGARET CORBITT<sup>1</sup>, KALINA GORNIK<sup>2</sup>, PAUL NICHOLSON<sup>1</sup> AND JAMES K. M. BROWN<sup>1,\*</sup>

<sup>1</sup>Department of Crop Genetics, John Innes Centre, Norwich Research Park, Norwich, NR4 7UH, UK

<sup>2</sup>Crop Protection Team, Crop and Soil Systems Group, SRUC, West Mains Road, Edinburgh, EH9 3JG, UK

## SUMMARY

*NAC* proteins are plant transcription factors that are involved in tolerance to abiotic and biotic stresses, as well as in many developmental processes. *Stress-responsive NAC1* (*SNAC1*) transcription factor is involved in drought tolerance in barley and rice, but has not been shown previously to have a role in disease resistance. Transgenic over-expression of *HvSNAC1* in barley cv. Golden Promise reduced the severity of *Ramularia* leaf spot (RLS), caused by the fungus *Ramularia collo-cygni*, but had no effect on disease symptoms caused by *Fusarium culmorum*, *Oculimacula yallundae* (eyespot), *Blumeria graminis* f. sp. *hordei* (powdery mildew) or *Magnaporthe oryzae* (blast). The *HvSNAC1* transcript was weakly induced in the RLS-susceptible cv. Golden Promise during the latter stages of *R. collo-cygni* symptom development when infected leaves were senescing. Potential mechanisms controlling *HvSNAC1*-mediated resistance to RLS were investigated. Gene expression analysis revealed no difference in the constitutive levels of antioxidant transcripts in either of the over-expression lines compared with cv. Golden Promise, nor was any difference in stomatal conductance or sensitivity to reactive oxygen species-induced cell death observed. Over-expression of *HvSNAC1* delayed dark-induced leaf senescence. It is proposed that mechanisms controlled by *HvSNAC1* that are involved in tolerance to abiotic stress and that inhibit senescence also confer resistance to *R. collo-cygni* and suppress RLS symptoms. This provides further evidence for an association between abiotic stress and senescence in barley and the development of RLS.

**Keywords:** biotroph, endophyte, hemibiotroph, necrotroph, plant–pathogen interaction, senescence, transgenic resistance.

## INTRODUCTION

Barley is the fourth most important cereal crop, grown in diverse environments worldwide (Newton *et al.*, 2011). Barley yields are threatened by a wide range of biotic stresses, such as pests and diseases (Walters *et al.*, 2012). As agricultural environments evolve through altered agronomic practices and climate change, crops such as barley are predicted to be at risk not only from current biotic stresses, but also from previously unrecognized pest and disease problems (Newton *et al.*, 2011; West *et al.*, 2012). Unfavourable environmental conditions caused by abiotic stresses, such as drought, flooding, extreme temperature, salinity and nutrient stress, can also cause yield losses that can be in excess of 50% (Bray *et al.*, 2000). To maintain food production, crop protection needs to guard against losses caused not only by recognised diseases and abiotic stresses, but also by new and emerging threats.

*Ramularia* leaf spot (RLS), caused by the ascomycete fungus *Ramularia collo-cygni*, is a newly important disease of barley across Europe (Walters *et al.*, 2008). The fungus is seed borne, but can also be spread by airborne spores that germinate on the leaf surface and penetrate through stomata (Havis *et al.*, 2014; Stabenheiner *et al.*, 2009). *Ramularia collo-cygni* lives as an asymptomatic endophyte during crop development, but can switch lifestyle late in the growing season to become a necrotrophic pathogen causing significant yield losses (Oxley and Havis, 2004; Walters *et al.*, 2008). The expression of disease symptoms is associated with an overall decline in the host antioxidant system (Schützendübel *et al.*, 2008) and progression of the disease results in an early onset of leaf senescence (Oxley *et al.*, 2008). However, the precise conditions that contribute to the fungal transition from endophyte to pathogen are not fully understood. Exposure of barley plants to abiotic stresses, such as high light, results in enhanced RLS symptom development, implying that plant stress may be important to the development of this disease (Makepeace *et al.*, 2008; Peraldi *et al.*, 2014). If abiotic stress factors are required to elicit the fungal transition from endophyte to pathogen, crops with

\*Correspondence: Email: graham.mcgrann@sruc.ac.uk; james.brown@jic.ac.uk

Present addresses:

†Crop Protection Team, Crop and Soil Systems Group, SRUC, West Mains Road, Edinburgh, EH9 3JG, UK.

‡RAGT Seeds Ltd, Grange Road, Ickleton, Essex, CB10 1TA, UK.

enhanced tolerance to abiotic stress may also be more resistant to RLS.

Abiotic and biotic stress factors elicit somewhat different plant responses, but some components involved in resistance to different types of stress may be shared (Atkinson *et al.*, 2013; Atkinson and Urwin, 2012; Narsai *et al.*, 2013). The identification and characterisation of genes that control tolerance to multiple abiotic and biotic stress factors could help to reveal the mechanisms that enable plants to cope with stress, as well as being potential candidates for use in plant breeding programmes. Transcription factors act as molecular switches involved in the regulation of plant developmental programmes and responses to diverse stresses. One of the largest families of transcription factors is the plant-specific NAC [no apical meristem (NAM), *Arabidopsis thaliana* transcription activation factor (ATAF1/2) and cup-shaped cotyledon (CUC2)] transcription factor superfamily. NAC transcription factors are involved in the regulation of many developmental processes, including secondary cell wall biosynthesis, senescence and biotic and abiotic stress tolerance (Puranik *et al.*, 2012). NAC genes can also promote and regulate reactive oxygen species (ROS) metabolism and homeostasis (Lee *et al.*, 2012; Wu *et al.*, 2012; You *et al.*, 2013). Functional diversification within the NAC protein family predates the separation of the monocot and dicot lineages (Nuruzzaman *et al.*, 2010) and many different NAC transcription factors have been characterised in cereals with various functions (Distelfeld *et al.*, 2012; Uauy *et al.*, 2006). Some cereal NAC transcription factors regulate responses to drought, cold and salinity stresses, whereas others act against pathogens (Chen *et al.*, 2013; Jensen *et al.*, 2007; Nakashima *et al.*, 2007; Sun *et al.*, 2013; Xia *et al.*, 2010). However, many NAC genes respond to both abiotic and biotic stresses, and the regulation of multiple stresses appears to be a common function of NAC transcription factors, indicating a degree of functional redundancy within this gene family (Nuruzzaman *et al.*, 2010).

NAC genes have been used in transgenic approaches to enhance abiotic stress tolerance in cereals. *Stress-responsive NAC1* (SNAC1) was first characterised in transgenic rice (*OsSNAC1*), demonstrating a positive role for this transcription factor in drought and salt tolerance (Hu *et al.*, 2006). *OsSNAC1* regulates abiotic and oxidative stress tolerance by enhancing the expression of stress-responsive genes (Hu *et al.*, 2006) and by targeting genes that control stomatal closure and ROS homeostasis (You *et al.*, 2013). Over-expression of the orthologous barley gene *HvSNAC1* also enhanced drought tolerance in barley plants (Al Abdallat *et al.*, 2014).

In this article, we demonstrate that SNAC1 has a previously unknown positive function in plant–pathogen interactions. Over-expression of *HvSNAC1* in barley reduces the severity of RLS symptoms as well as fungal colonisation by *R. collo-cygni*. This appears to be related to the delayed senescence of *HvSNAC1*

over-expression lines rather than alterations in antioxidant capacity or sensitivity to ROS. The specificity of the response to RLS and the link between *HvSNAC1* over-expression and enhanced leaf longevity provide further insights into the involvement of senescence in the interaction between barley and RLS.

## RESULTS

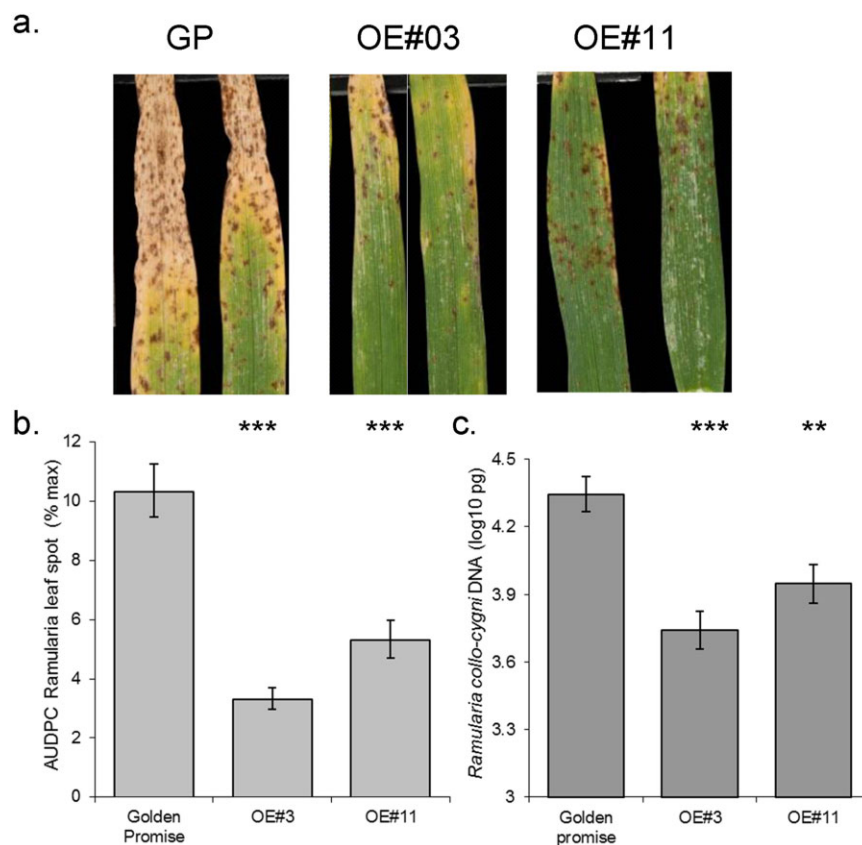
### Disease development on *HvSNAC1* over-expression lines

The role of *HvSNAC1* in defence against fungal diseases was explored using two independent transgenic barley cv. Golden Promise (GP) lines, OE#3 and OE#11, which both exhibited consistently elevated *HvSNAC1* transcript levels in seedling prophyll leaves (Fig. S1, see Supporting Information). Disease symptom expression was assessed on plants inoculated with *R. collo-cygni*, as well as the hemibiotrophic fungal pathogens *Oculimacula yallundae*, *Magnaporthe oryzae* and *Fusarium culmorum*, which differ in the duration of biotrophic growth before entering necrotrophic development, ranging from weeks to days, and the obligate biotroph *Blumeria graminis* f. sp. *hordei*. Typical development of *R. collo-cygni* was observed on cv. GP with the wild-type *HvSNAC1* gene, a barley variety that shows a moderate degree of susceptibility to isolate Rcc09B4. RLS first became visible from 10 to 12 days post-inoculation (dpi). The number of spots increased over time with lesions coalescing as the leaf began to senesce (Fig. 1a). The development of *R. collo-cygni* symptoms was reduced in both transgenic lines compared with GP ( $P < 0.001$ ; Fig. 1a,b). Significantly smaller amounts of *R. collo-cygni* DNA were recorded in both transgenic lines compared with GP (60%–75% lower than GP; Fig. 1c). No significant differences were observed in *R. collo-cygni* DNA levels in seeds of GP and the two transgenic lines that were used in the inoculation experiments. ( $P = 0.4$ ; results not shown).

No significant differences were observed in the size of the lesions formed by *F. culmorum* (Fig. 2a;  $P = 0.2$ ) or the number of lesions formed by *M. oryzae* (Fig. 2b;  $P = 0.3$ ) on the leaves of either transgenic line compared with GP. Over-expression of *HvSNAC1* also had no significant effect on disease symptom development of the stem base eyespot disease caused by *O. yallundae* (Fig. 2c;  $P = 0.3$ ) or on colony formation by *B. graminis* f. sp. *hordei* (Fig. 2d;  $P = 0.8$ ).

### Expression analysis of *HvSNAC1* transcript following *R. collo-cygni* inoculation

Time course analysis of *HvSNAC1* transcript expression during *R. collo-cygni* infection in GP leaves indicated that *HvSNAC1* was not differentially regulated during the early asymptomatic stages of the *R. collo-cygni* infection process, up to 10 dpi (Fig. 3). A small



**Fig. 1** Development of *Ramularia* leaf spot (RLS) in transgenic *HvSNAC1* over-expression (OE) barley lines. (a) Disease symptom development at 21 days post-inoculation (dpi) on Golden Promise (GP), OE#3 and OE#11. (b) Area under the disease progress curve (AUDPC) of RLS. (c) *Ramularia collo-cygni* DNA in leaves of transgenic and wild-type plants at 21 dpi. Error bars indicate  $\pm 1$ SE. \*\*\* $P < 0.001$  and \*\* $P < 0.01$  for comparison of means of OE lines with GP.

increase in *HvSNAC1* transcript level was observed at the later stages of infection when leaves exhibited severe symptoms and had begun to senesce (Fig. 1a and 3).

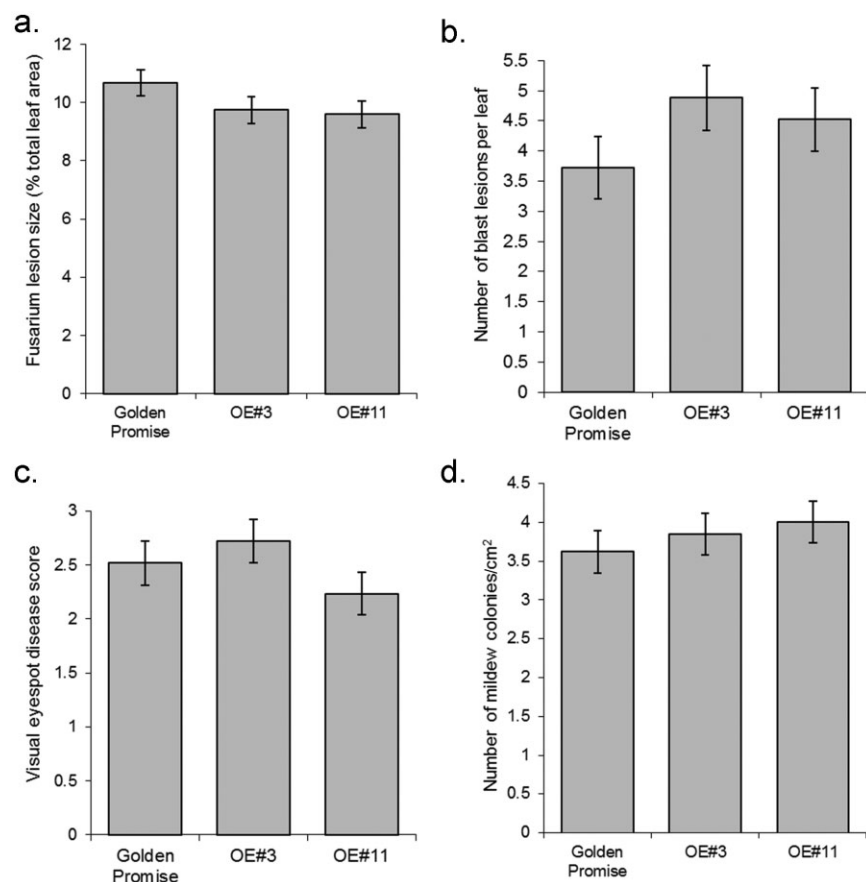
#### Effect of *HvSNAC1* over-expression on leaf antioxidant transcript levels and sensitivity to ROS-induced cell death

The onset of RLS symptoms in the field has been associated with an overall decline in the host antioxidant system (Schützendübel *et al.*, 2008). Transcript profiling of the major barley ROS scavengers, including catalases (CAT1 + CAT2), ascorbate peroxidases (APX1 + APX2), glutathione peroxidases (GPX1 + GPX2), glutathione reductase (GR1) and copper/zinc superoxide dismutase (CSD1), was used to assess the effect of *HvSNAC1* over-expression on the antioxidant system of barley. Constitutive gene expression levels of all of the antioxidant transcripts tested were similar in both transgenic lines to those in GP (Fig. 4a). The role of *HvSNAC1* over-expression on ROS-induced cell death was examined by testing the effect of different ROS donors on cell death lesion formation in lines OE#3 and OE#11 relative to GP. No significant differences in lesion size produced by the hydrogen peroxide donor alloxan ( $P = 0.9$ ), the mitochondrial superoxide donor menadione ( $P = 0.7$ ) and the chloroplastic superoxide donor

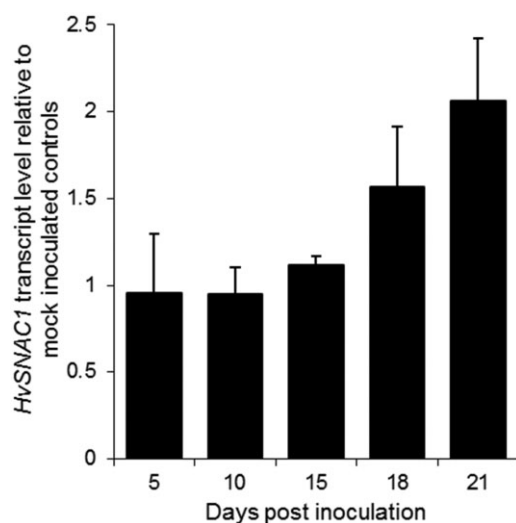
methyl viologen ( $P = 0.1$ ) were observed between either of the transgenic lines and GP (Fig. 4b).

#### Effect of *HvSNAC1* over-expression on leaf senescence and stomatal conductance

In the field, RLS symptoms become visible during the later developmental stages of the host as the plant begins to flower and the leaves start to senesce (Schützendübel *et al.*, 2008). It is not currently known whether leaf senescence is a cause or consequence of disease. Dark-induced senescence, a model system used to study leaf senescence in plants (Gan and Amasino, 1997), was used to test the effect of over-expression of *HvSNAC1* on leaf senescence. All three lines showed a decline in relative chlorophyll content over time (Fig. 5a,  $P < 0.001$ ). There was no significant difference between the relative chlorophyll contents of GP and the two transgenic lines at day 0 or after 2 days of dark treatment. From day 4 onwards, there was a significantly slower decline in relative chlorophyll content in both over-expression lines OE#3 and OE#11 compared with GP ( $P < 0.001$ ). This slower decline in relative chlorophyll content was also observed at day 6 ( $P < 0.001$ ) and day 8 ( $P < 0.01$ ) in both transgenic lines, indicating that over-expression of *HvSNAC1* delays dark-induced leaf senescence (Fig. 5a).



**Fig. 2** Development of disease symptoms caused by *Fusarium culmorum* (a), *Magnaporthe oryzae* (b), *Oculimacula yallundae* (c) and *Blumeria graminis* f. sp. *hordei* (d) in transgenic *HvSNAC1* over-expression (OE) barley lines. Error bars indicate  $\pm 1SE$ . None of the mean scores of the OE lines were significantly different from Golden Promise (GP) ( $P > 0.05$ ).



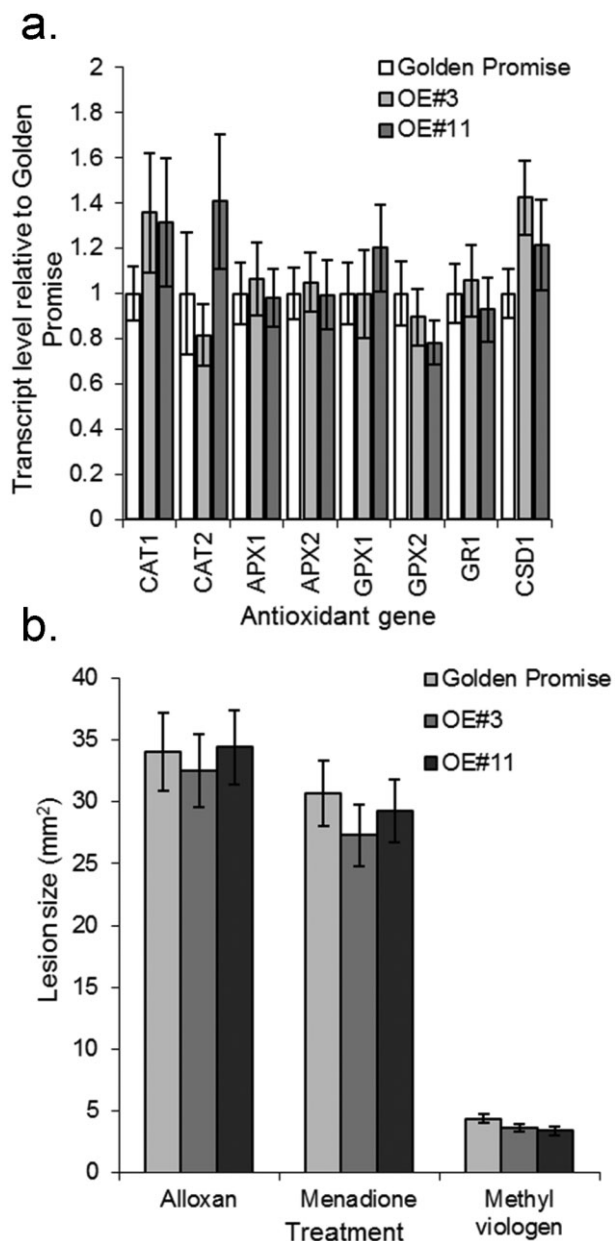
**Fig. 3** Quantitative reverse transcription-polymerase chain reaction (qRT-PCR) time course analysis of *HvSNAC1* transcript expression in wild-type Golden Promise leaves following infection with *Ramularia collo-cygni*. Transcript levels are presented relative to mock-inoculated controls. Error bars indicate SE.

Given the role of *HvSNAC1* in drought tolerance and that *R. collo-cygni* infects leaves through the stomata, the effect of the over-expression of *HvSNAC1* on stomatal closure was examined. No significant differences in stomatal conductance between the transgenic lines OE#3 and OE#11 and GP were observed (Fig. 5b,  $P = 0.7$ ).

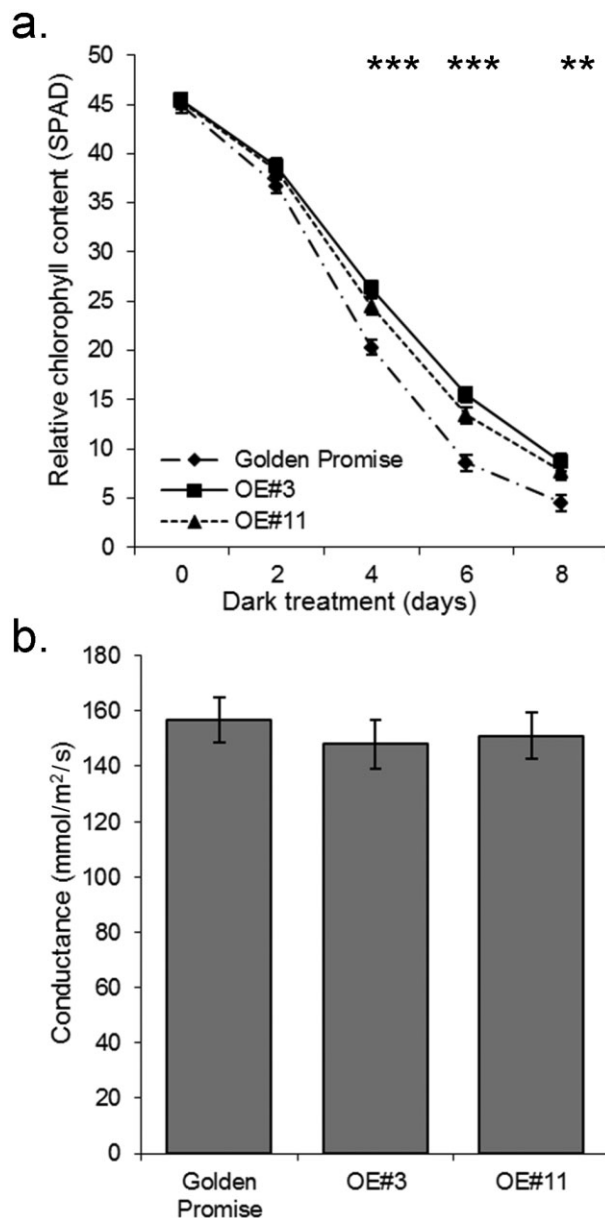
## DISCUSSION

As a recently emerging problem of barley, the factors that result in the switching of the endophytic fungus *R. collo-cygni* to a necrotrophic pathogen, resulting in disease, are not fully understood. The development of RLS symptoms typically occurs late in the growing season and appears to be associated with the action of environmental stresses on the host (Makepeace *et al.*, 2008; Oxley and Havis, 2004; Peraldi *et al.*, 2014; Schützendübel *et al.*, 2008). NAC transcription factors act as central modulators of stress responses that function as transcriptional activators to regulate the expression of stress-related genes, including those involved in defence, detoxification and redox homeostasis (Hu *et al.*, 2006; Nakashima *et al.*, 2007; Sun *et al.*, 2013). *SNAC1* transcription factors enhance the drought tolerance of cereals (Al





**Fig. 4** Effect of *HvSNAC1* over-expression on barley redox system. (a) Quantitative reverse transcription-polymerase chain reaction (qRT-PCR) analysis of the constitutive transcript levels of the major reactive oxygen species scavengers in *HvSNAC1* over-expression lines. Transcript levels are presented relative to wild-type Golden Promise leaves. APX1, ascorbate peroxidase 1; APX2, ascorbate peroxidase 2; CAT1, catalase 1; CAT2, catalase 2; CSD1, copper/zinc superoxide dismutase 1; GPX1, glutathione peroxidase 1; GPX2, glutathione peroxidase 2; GR1, glutathione reductase 1. Error bars indicate SE (b) Lesion development caused by the reactive oxygen species donors alloxan, menadione and methyl viologen in transgenic *HvSNAC1* over-expression barley lines. Error bars indicate  $\pm 1$ SE.



**Fig. 5** Effect of *HvSNAC1* over-expression (OE) on dark-induced senescence (a) and stomatal closure (b) compared with Golden Promise (GP). Error bars indicate  $\pm 1$ SE. \*\*\* $P < 0.001$  and \*\* $P < 0.01$  for comparison of means of OE lines with GP.

Abdallat *et al.*, 2014; Hu *et al.*, 2006). Here, we show that over-expression of the barley *SNAC1* gene confers a small increase in resistance to RLS, but has no effect on the interaction between barley and the fungal pathogens *B. graminis* f. sp. *hordei*, *F. culmorum*, *M. oryzae* and *O. yallundae*. This implies that there is a specific interaction between the endophytic parasite *R. collo-cygni* and *SNAC1*.

Symbiotic interactions between fungal endophytes and plants are frequently benign or beneficial to the host. However, when host conditions become unfavourable for an endophytic lifestyle, these fungi can become pathogenic (Rodriguez and Redman, 2008). The development of RLS symptoms typically occurs following conditions that are expected to alter host ROS status (Makepeace *et al.*, 2008; Peraldi *et al.*, 2014; Schützendübel *et al.*, 2008). Increased cellular ROS concentrations, if not controlled, can result in cell death, which promotes the development of necrotrophic pathogens (Heller and Tudzynski, 2011). NAC genes regulate ROS levels in plants under stressful conditions and the rice orthologue of *HvSNAC1*, *OsSNAC1*, regulates ROS homeostasis through interactions with genes, such as *OsSRO1* (You *et al.*, 2013). Over-expression of *HvSNAC1* did not influence directly the regulation of ROS scavenger transcript expression or sensitivity to ROS-induced cell death (Fig. 4). This implies that the tolerance of transgenic plants over-expressing *HvSNAC1* to RLS and restriction of the necrotrophic development of the fungus do not result from elevated resistance to ROS-related damage.

*Ramularia collo-cygni* is transmitted by infected seed (Havis *et al.*, 2014) and spore-borne foliar infections that occur through stomatal openings (Stabentheiner *et al.*, 2009). Seed-borne inoculum contributes to the initial infection, whereas spore-borne infections later in the season may play a role in secondary infections and fungal dissemination (Havis *et al.*, 2014). The relative contribution of infection via both of these modes of transmission towards final disease levels is not fully understood. Stomata form points of entry for the fungus and, following spore or mycelium inoculation, *R. collo-cygni* grows intercellularly with conidiophores emerging from stomata (Stabentheiner *et al.*, 2009; Thirugnanasambandam *et al.*, 2011). The rice orthologue of *HvSNAC1*, *OsSNAC1*, can regulate stomatal closure (Hu *et al.*, 2006) through its action on *OsSRO1* (You *et al.*, 2013), but we found no effect of *HvSNAC1* over-expression in barley on constitutive levels of stomatal closure (Fig. 5b), corroborating the work of Al Abdallat *et al.* (2014). These data suggest that the reduced susceptibility of the *HvSNAC1* over-expression lines to *R. collo-cygni* is unlikely to be a result of fewer infection events and reduced fungal penetration resulting from more tightly closed stomata. The effect of *HvSNAC1* over-expression specifically on seed-borne RLS epidemiology remains to be determined.

Disease development occurs late in the growing season and has been associated with crop senescence (Oxley *et al.*, 2008; Schützendübel *et al.*, 2008). Over-expression of *HvSNAC1* delayed dark-induced leaf senescence in both transgenic lines (Fig. 5a). *HvSNAC1* transcript levels showed a weak induction in GP during the later stages of RLS symptom development (Fig. 3). However, *HvSNAC1* transcripts are induced in senescing flag leaves (Christiansen *et al.*, 2011), and so it is unclear whether the increased expression of this gene is a response to stress caused by fungal colonisation or is a consequence of premature leaf senes-

cence associated with the later stages of disease development (Fig. 1a; Oxley *et al.*, 2008; Walters *et al.*, 2008). NAC transcription factors can positively or negatively affect senescence (Distelfeld *et al.*, 2012; Lee *et al.*, 2012; Uauy *et al.*, 2006; Wu *et al.*, 2012; Zhou *et al.*, 2013). *TtNAM-B1* and *OsNAP*, which are related to the *SNAC* subgroup of cereal NAC genes (Nuruzzaman *et al.*, 2010), promote leaf senescence in wheat and rice (Uauy *et al.*, 2006; Zhou *et al.*, 2013), respectively, whereas *HvSNAC1* delays the process in barley (Fig. 5a). The results reported here imply that the over-expression of a gene which suppresses senescence in barley also suppresses the growth of *R. collo-cygni* and the development of RLS symptoms. NAC transcription factors regulate senescence through different processes, including manipulation of ROS and hormone pathways. Positive regulation of senescence by *OsNAP* appears to be associated with jasmonate biosynthesis and signalling pathways (Zhou *et al.*, 2013), but the relationship between *SNAC1* and plant hormones is unknown. Whether the effect of *SNAC1* on leaf senescence operates through previously characterised or novel hormone signalling pathways remains to be revealed.

The longevity of green leaf area has been genetically linked to drought tolerance in cereals (Foulkes *et al.*, 2007). NAC gene mutants that delay senescence in the model plant *Arabidopsis thaliana* also show enhanced tolerance to abiotic stresses, implying an association between senescence and stress (Lee *et al.*, 2012; Wu *et al.*, 2012). In the field, there is some evidence that leaf senescence promotes RLS (Schützendübel *et al.*, 2008), and barley varieties that are highly susceptible to RLS tend to senesce early as a result of the disease (Oxley *et al.*, 2008). Although *HvSNAC1* over-expression lines, which have reduced susceptibility to RLS (Fig. 1), have a small effect on reducing the rate of leaf senescence (Fig. 5), further experiments are required to elucidate the interactions of the signalling pathways regulated by *HvSNAC1* with *R. collo-cygni*, stress and senescence.

Deciphering the signalling networks controlled by *HvSNAC1* will further our understanding of the host processes that contribute to the expression of RLS. *HvSNAC1* over-expression affected the development of RLS, but had no significant effect on four other diseases (Fig. 2), despite the fact that the *HvSNAC1* transcript was regulated upwards and downwards by infection with *Fusarium* and *B. graminis*, respectively (Al Abdallat *et al.*, 2014). Therefore, it appears that *HvSNAC1* has a specific effect on RLS and is not a major factor contributing to the interaction between barley and other diseases (Fig. 2). The association between symptom development of this late season disease, plant stress and senescence implies that the delayed senescence phenotype of the *HvSNAC1* over-expression lines is involved in the reduced growth of *R. collo-cygni* and expression of RLS symptoms. As a newly important pathogen, little is known about the host genetic components involved in the interaction with *R. collo-cygni*. Recent evidence that mutant *mlo* alleles, which confer resistance to the biotrophic

powdery mildew fungus, increase susceptibility to RLS (McGrann *et al.*, 2014) further implies that senescence is involved in the aetiology of RLS. The wild-type *MLO* gene responds to both biotic and abiotic stresses (Baker *et al.*, 2000) and mutant *mlo* alleles accelerate the rate of leaf senescence once the process has begun (Piffanelli *et al.*, 2002). Whether the *mlo*-enhanced susceptibility to RLS is specifically linked to senescence or to one or more of the other pleiotropic effects caused by the *mlo* mutation requires further experimentation. Interpretation of the link between RLS, senescence and stress responses may provide insights into why RLS has recently emerged as an important disease of barley.

## EXPERIMENTAL PROCEDURES

### Plant material

Two transgenic lines, OE#3 and OE#11, from independent transformation events over-expressing *HvSNAC1* (*HvNAC003*; Christiansen *et al.*, 2011) were used in this study (Al Abdallat *et al.*, 2014). Seeds were sown in  $8 \times 8 \times 10\text{-cm}^3$  pots in Levington F2 compost medium (Scotts Professional, Ipswich, UK), and grown under a 16 h/8 h day/night photoperiod at day/night temperatures of 18/12 °C provided by 220  $\mu\text{mol/m}^2/\text{s}$  fluorescent lighting in a controlled environment room (Sanyo Gallenkamp PLC, Loughborough, UK).

### Pathogen inoculations

*Ramularia collo-cygni* isolate Rcc09B4 was used to inoculate barley plants following the protocol of Makepeace *et al.* (2008) with the modifications of Peraldi *et al.* (2014). RLS development was scored as the percentage leaf area covered with disease symptoms three to five times over 21 dpi and the area under the disease progress curve (AUDPC) was calculated. Three independent replicate experiments, each containing an individual  $8 \times 8 \times 10\text{-cm}^3$  pot of 10 seeds of each line, were inoculated with *R. collo-cygni*. *Oculimacula yallundae* isolate P149 was inoculated at the stem base of 21-day-old plants using the method of Chapman *et al.* (2008). A single experiment was inoculated in a randomised block design, consisting of five blocks, with each block containing five plants of each line. The penetration of stem bases by *O. yallundae* was assessed 8 weeks post-inoculation using the scale of Scott (1971). *Magnaporthe oryzae* isolate BR32 was inoculated at a concentration of  $10^5$  spores/mL in two independent experiments, as described previously (Tufan *et al.*, 2009). In each experiment, 10 seeds of each line were sown in individual  $8 \times 8 \times 10\text{-cm}^3$  pots and disease was assessed at 6 dpi as the number of blast lesions present on the second seedling leaf of each plant. The development of *F. culmorum* isolate Fu42 was assessed on detached leaves inoculated with two 5- $\mu\text{L}$  droplets of  $10^6$  conidia/mL amended with 75  $\mu\text{M}$  deoxynivalenol (Chen *et al.*, 2009). Photographs of disease lesion development were taken 48 h post-inoculation and the lesion area was measured using ImageJ software (Abramoff *et al.*, 2004). Two independent replicate experiments, each containing a minimum of six replicate leaves of each line, were inoculated. *Blumeria graminis* f. sp. *hordei* isolate CC148 was used to inoculate leaf segments from 14-day-old prophyll leaves

following the method of Boyd *et al.* (1994). Disease development was assessed at 14 dpi as the number of colonies observed per square centimetre of leaf area in four independent replicate experiments, each consisting of a minimum of eight replicate leaves of each line.

The response of *HvSNAC1* over-expression lines to each pathogen was analysed separately using a general linear model (GLM) in GenStat v.15 (Payne *et al.*, 2009). Each GLM assessed the variation attributable to experimental replicate, block and line, and calculated the predicted mean disease scores on each line. Predicted means were used to compare the disease development of each pathogen on the two transgenic lines and GP with *t*-tests using the residual standard error of the respective model.

### Quantitative reverse transcription-polymerase chain reaction (qRT-PCR) gene expression assays

Constitutive levels of *HvSNAC1* and ROS scavenger gene transcripts were assessed in each of the two transgenic lines compared with GP using qRT-PCR. RNA was extracted from 14-day-old prophyll leaves, processed and converted to cDNA (Colebrook *et al.*, 2012). Transcripts were amplified with gene-specific primers (Table S1, see Supporting Information; Shagimardanova *et al.*, 2010) and the Sybr Green JumpStart™ Taq Ready mix system (Sigma-Aldrich, St. Louis, MO, USA), as described previously (Colebrook *et al.*, 2012). cDNA samples were normalised with geNorm (Vandesompele *et al.*, 2002) using five reference genes (elongation factor 1 $\alpha$ , glyceraldehyde 3-phosphate dehydrogenase, cyclophilin, ubiquitin,  $\alpha$ -tubulin; Burton *et al.*, 2004; McGrann *et al.*, 2009; Rostoks *et al.*, 2003). Data were collected from three independent replicate experiments, each consisting of at least two independent samples of each line.

Expression of the *HvSNAC1* transcript during the host response to *R. collo-cygni* infection was assessed in GP. Seedlings were inoculated as described above with either Rcc09B4 or potato dextrose broth containing no fungus as the control. Two samples, each consisting of two pooled prophyll leaves from Rcc09B4 and control inoculated plants, were collected at 5, 10, 15, 18 and 21 dpi. Samples were processed as above and the expression of the *HvSNAC1* transcript was compared between Rcc09B4 and control inoculated samples at each time point. Data were obtained from three independent replicate experiments.

### qPCR quantification of *R. collo-cygni* DNA levels in planta

In order to estimate the amount of *R. collo-cygni* within host leaves, fungal DNA levels were measured by qPCR (Taylor *et al.*, 2010). DNA was extracted from five leaf samples of each line collected at 21 dpi using the DNeasy Plant Mini Kit (Qiagen, Hilden, Germany). Fungal DNA levels in three independent replicate experiments were quantified by qPCR using a CFX96 thermocycler (Bio-Rad, Hercules, CA, USA). Differences in  $\log_{10}$  transformed fungal DNA levels between the two transgenic lines and GP were analysed using a GLM in GenStat v.15 with experimental replicate and line as factors. *Ramularia collo-cygni* DNA levels in seeds were assessed using the methods of Havis *et al.* (2014).

### ROS-induced cell death assays

ROS-induced cell death assays were performed on barley leaves following the method of Saville (2011). Prophyll leaves from 14-day-old plants of



each transgenic line and GP were detached and suspended across agar (1% agar w/v supplemented with 100 mg/L benzimidazole; Sigma-Aldrich) bridges in clear plastic boxes. The sensitivity of each line to ROS-induced cell death was tested by adding 2 µL of solution, supplemented with 0.5% v/v Tween20, of the following ROS donors to the centre of each leaf: 200 mM alloxan (Sigma-Aldrich), 100 mM menadione (Sigma-Aldrich) and 25 µM methyl viologen (Sigma-Aldrich). Inoculated leaves were stored under constant light (15–20 µmol/m<sup>2</sup>/s) for 96 h at room temperature. After incubation, each box was photographed and the lesion size was measured using ImageJ. ROS-induced lesion formation was measured from three independent replicate experiments, each consisting of a minimum of 24 replicate leaves of each line for each ROS donor. General linear modelling was used to estimate the effects of replicate experiment and line. Each ROS donor was analysed separately.

### Stomatal conductance measurements

The effect of over-expression of *HvSNAC1* on stomatal conductance was measured using an AP4 cycling porometer (Delta-T Devices Ltd, Cambridge, UK) based on the methods described by Prats *et al.* (2006). Leaf water conductance measurements were taken from the prophyll leaf of 5–10 14-day-old seedlings of each line. The experiment was performed four times. Data were analysed using a GLM in GenStat v.15 with experiment and line as factors.

### Dark-induced senescence assay

Dark-induced senescence in barley leaves was measured following the method outlined by Peraldi (2012). Prophyll leaves from 14-day-old seedlings were excised and an indirect measurement of leaf chlorophyll content was taken using a SPAD 502 Plus Chlorophyll Meter (Konica Minolta, Warrington, UK). Three measurements were taken across each leaf to cover the distal, middle and basal portions, and averaged. After the first measurement (day 0), leaves were placed on damp tissue paper in 10-cm<sup>2</sup> plastic Petri dishes, covered with aluminium foil and kept in the dark at room temperature to induce senescence. Further measurements were taken on each leaf after 2, 4, 6 and 8 days of dark treatment. Three replicate experiments were performed with measurements taken from six leaves of each line in each experiment. Linear mixed modelling of repeated measurements was used to evaluate differences in SPAD readings between each line at the different time points employing the uniform correlation/split plot in time covariance matrix. Fixed factors included day, experiment, line and the interactions between day and line, and experiment and day. The random factor was the leaf-by-day interaction term. Significant differences between lines at different days were subsequently assessed using a *t*-test.

### ACKNOWLEDGEMENTS

We thank Ayed Al-Abdallat (University of Jordan) and Wendy Harwood John Innes Centre (JIC) for seeds of the *HvSNAC1* transgenic lines, and Mike Grimmer (ADAS) for the loan of the AP4 cycling porometer. We also thank Didier Tharreau (CIRAD, Montpellier) for providing the *Magnaporthe oryzae* isolate BR32, which was used under Defra Plant Health Licence 51098/198798/1 (3/2013), and Neil Havis (SRUC) for useful discussions on

the biology of *Ramularia collo-cygni*. This research was supported by the Biotechnology and Biological Sciences Research Council, RESAS, AHDB-HGCA and a consortium of 11 companies (BASF, Bayer, KWS, Lantmännen, Limagrain, LS, NIAB-TAG, Saaten-Union, Secobra, Sejet, Syngenta) through Sustainable Arable LINK, by grant BB/G024006/1 from BBSRC and by the John Innes Foundation. SRUC receives grant-in-aid from the Scottish Government. CL was funded by a student bursary from the British Society for Plant Pathology.

### REFERENCES

- Abramoff, M.D., Magalhaes, P.J. and Ram, S.J. (2004) Image processing with ImageJ. *Biophotonics Int.* **11**, 36–42.
- Al Abdallat, A.M., Ayad, J.Y., Abu Elenein, J.M., Al Ajlouni, Z. and Harwood, W.A. (2014) Overexpression of the transcription factor *HvSNAC1* improves drought tolerance in barley (*Hordeum vulgare* L.). *Mol. Breed.* **33**, 401–414.
- Atkinson, N.J. and Urwin, P.E. (2012) The interaction of plant biotic and abiotic stresses: from genes to the field. *J. Exp. Bot.* **63**, 3523–3543.
- Atkinson, N.J., Lilley, C.J. and Urwin, P.E. (2013) Identification of genes involved in the response of *Arabidopsis* to simultaneous biotic and abiotic stresses. *Plant Physiol.* **162**, 2028–2041.
- Baker, S.J., Newton, A.C. and Gurr, S.J. (2000) Cellular characteristics of temporary partial breakdown of *mlo*-resistance in barley to powdery mildew. *Physiol. Mol. Plant Pathol.* **56**, 1–11.
- Boyd, L.A., Smith, P.H., Green, R.M. and Brown, J.K.M. (1994) The relationship between the expression of defense-related genes and mildew development in barley. *Mol. Plant-Microbe Interact.* **7**, 401–410.
- Bray, E.A., Bailey-Serres, J. and Weretilnyk, E. (2000) Response to abiotic stresses. In: *Biochemistry and Molecular Biology of Plants* (Gruissem, W., Buchanan, B. and Jones, R., eds), pp. 1158–1203. Rockville, MD: American Society of Plant Physiologists.
- Burton, R.A., Shirley, N.J., King, B.J., Harvey, A.J. and Fincher, G.B. (2004) The *CesA* gene family of barley. Quantitative analysis of transcripts reveals two groups of co-expressed genes. *Plant Physiol.* **134**, 224–236.
- Chapman, N.H., Burt, C., Dong, H. and Nicholson, P. (2008) The development of PCR-based markers for the selection of eyespot resistance genes *Pch1* and *Pch2*. *Theor. Appl. Genet.* **117**, 425–433.
- Chen, X., Steed, A., Travella, S., Keller, B. and Nicholson, P. (2009) *Fusarium graminearum* exploits ethylene signalling to colonize dicotyledonous and monocotyledonous plants. *New Phytol.* **182**, 975–983.
- Chen, Y.-J., Perera, V., Christiansen, M.W., Holme, I.B., Gregersen, P.L., Grant, M.R., Collinge, D.B. and Lyngkjær, M.F. (2013) The barley *HvNAC6* transcription factor affects ABA accumulation and promotes basal resistance against powdery mildew. *Plant Mol. Biol.* **83**, 577–590.
- Christiansen, M.W., Holm, P.B. and Gregersen, P.L. (2011) Characterization of barley (*Hordeum vulgare* L.) NAC transcription factors suggests conserved functions compared to both monocots and dicots. *BMC Res. Notes*, **4**, 302.
- Colebrook, E.H., Creissen, G., McGrann, G.R.D., Dreos, R., Lamb, C. and Boyd, L.A. (2012) Broad-spectrum acquired resistance in barley induced by the *Pseudomonas* pathosystem shares transcriptional components with *Arabidopsis* systemic acquired resistance. *Mol. Plant-Microbe Interact.* **25**, 658–667.
- Distelfeld, A., Pearce, S.P., Avni, R., Scherer, B., Uauy, C., Piston, F., Slade, A., Zhao, R. and Dubcovsky, J. (2012) Divergent functions of orthologous NAC transcription factors in wheat and rice. *Plant Mol. Biol.* **78**, 515–524.
- Foulkes, M.J., Sylvester-Bradley, R., Weightman, R. and Snape, J.W. (2007) Identifying physiological traits associated with improved drought resistance in winter wheat. *Field Crops Res.* **103**, 11–24.
- Gan, S.S. and Amasino, R.M. (1997) Making sense of senescence—molecular genetic regulation and manipulation of leaf senescence. *Plant Physiol.* **113**, 313–319.
- Havis, N.D., Nyman, N. and Oxley, S.J.P. (2014) Evidence for seed transmission and asymptomatic growth of *Ramularia collo-cygni* in barley (*Hordeum vulgare*). *Plant Pathol.* **63**, 929–936. doi: 10.1111/ppa.12162.
- Heller, J. and Tudzynski, P. (2011) Reactive oxygen species in phytopathogenic fungi: signaling, development, and disease. *Annu. Rev. Phytopathol.* **49**, 369–390.
- Hu, H., Dai, M., Yao, J., Xiao, B., Li, X., Zhang, Q. and Xiong, L. (2006) Overexpressing a NAM, ATAF, and CUC (NAC) transcription factor enhances drought resistance and salt tolerance in rice. *Proc. Natl. Acad. Sci. USA*, **103**, 12 987–12 992.
- Jensen, M.K., Rung, J.H., Gregersen, P.L., Gjetting, T., Fuglsang, A.T., Hansen, M., Joehnk, N., Lyngkjær, M.F. and Collinge, D.B. (2007) The *HvNAC6* transcription

- factor: a positive regulator of penetration resistance in barley and *Arabidopsis*. *Plant Mol. Biol.* **65**, 137–150.
- Lee, S., Seo, P.J., Lee, H.-J. and Park, C.-M. (2012) A NAC transcription factor NTL4 promotes reactive oxygen species production during drought-induced leaf senescence in *Arabidopsis*. *Plant J.* **70**, 831–844.
- Makepeace, J.C., Havis, N.D., Burke, J.I., Oxley, S.J.P. and Brown, J.K.M. (2008) A method of inoculating barley seedlings with *Ramularia collo-cygni*. *Plant Pathol.* **57**, 991–999.
- McGrann, G.R.D., Townsend, B.J., Antoniw, J.F., Asher, M.J.C. and Mutasa-Goettgens, E.S. (2009) Barley elicits a similar early basal defence response during host and non-host interactions with *Polymyxa* root parasites. *Eur. J. Plant Pathol.* **123**, 5–15.
- McGrann, G.R.D., Stavrinos, A., Russell, J., Corbitt, M.M., Booth, A., Chartrain, L., Thomas, W.T.B. and Brown, J.K.M. (2014) A trade-off between *mlo* resistance to powdery mildew and increased susceptibility of barley to a newly important disease, *Ramularia* leaf spot. *J. Exp. Bot.* **65**, 1025–1037.
- Nakashima, K., Tran, L.-S.P., Van Nguyen, D., Fujita, M., Maruyama, K., Todaka, D., Ito, Y., Hayashi, N., Shinozaki, K. and Yamaguchi-Shinozaki, K. (2007) Functional analysis of a NAC-type transcription factor OsNAC6 involved in abiotic and biotic stress-responsive gene expression in rice. *Plant J.* **51**, 617–630.
- Narsai, R., Wang, C., Chen, J., Wu, J., Shou, H. and Whelan, J. (2013) Antagonistic, overlapping and distinct responses to biotic stress in rice (*Oryza sativa*) and interactions with abiotic stress. *BMC Genomics*, **14**, 93.
- Newton, A.C., Flavell, A.J., George, T.S., Leat, P., Mullholland, B., Ramsay, L., Revored-Giha, C., Russell, J., Steffenson, B.J., Swanson, J.S., Thomas, W.T.B., Waugh, R., White, P.J. and Bingham, I.J. (2011) Crops that feed the world 4. Barley: a resilient crop? Strengths and weaknesses in the context of food security. *Food Secur.* **3**, 141–178.
- Nuruzzaman, M., Manimekalai, R., Sharoni, A.M., Satoh, K., Kondoh, H., Ooka, H. and Kikuchi, S. (2010) Genome-wide analysis of NAC transcription factor family in rice. *Gene*, **465**, 30–44.
- Oxley, S.J.P. and Havis, N.D. (2004) Development of *Ramularia collo-cygni* on spring barley and its impact on yield. *Crop Protection in Northern Britain*, **2004**, 147–152.
- Oxley, S.J.P., Havis, N.D., Brown, J.K.M., Makepeace, J.C. and Fountaine, J. (2008) Impact and interactions of *Ramularia collo-cygni* and oxidative stress in barley. *HGCA Project Report No. 431*. London, UK: HGCA Publications.
- Payne, R.W., Murray, D.A., Harding, S.A., Baird, D.B. and Soutar, D.M. (2009) *GenStat for Windows*, 12th edn. Introduction. Hemel Hempstead, Hertfordshire: VSN International.
- Peraldi, A. (2012) *Brachypodium distachyon* as a genetic model pathosystem to study resistance against fungal pathogens of small grain cereals. PhD thesis, University of East Anglia, Norwich.
- Peraldi, A., Griffe, L.L., Burt, C., McGrann, G.R.D. and Nicholson, P. (2014) *Brachypodium distachyon* exhibits compatible interactions with *Oculimacula* spp. and *Ramularia collo-cygni*, providing the first pathosystem model to study eyespot and ramularia leaf spot diseases. *Plant Pathol.* **63**, 554–562.
- Piffanelli, P., Zhou, F.S., Casais, C., Orme, J., Jarosch, B., Schaffrath, U., Collins, N.C., Panstruga, R. and Schulze-Lefert, P. (2002) The barley MLO modulator of defense and cell death is responsive to biotic and abiotic stress stimuli. *Plant Physiol.* **129**, 1076–1085.
- Prats, E., Gay, A.P., Mur, L.A.J., Thomas, B.J. and Carver, T.L.W. (2006) Stomatal lock-open, a consequence of epidermal cell death, follows transient suppression of stomatal opening in barley attacked by *Blumeria graminis*. *J. Exp. Bot.* **57**, 2211–2226.
- Puranik, S., Sahu, P.P., Srivastava, P.S. and Prasad, M. (2012) NAC proteins: regulation and role in stress tolerance. *Trends Plant Sci.* **17**, 369–381.
- Rodríguez, R. and Redman, R. (2008) More than 400 million years of evolution and some plants still can't make it on their own: plant stress tolerance via fungal symbiosis. *J. Exp. Bot.* **59**, 1109–1114.
- Rostoks, N., Schmierer, D., Kudrna, D. and Kleinhofs, A. (2003) Barley putative hypersensitive induced reaction genes: genetic mapping, sequence analyses and differential expression in disease lesion mimic mutants. *Theor. Appl. Genet.* **107**, 1094–1101.
- Saville, R.J. (2011) Understanding DELLA in wheat linking genotype to phenotype. PhD thesis, University of East Anglia, Norwich.
- Schützendübel, A., Stadler, M., Wallner, D. and von Tiedemann, A. (2008) A hypothesis on physiological alterations during plant ontogenesis governing susceptibility of winter barley to ramularia leaf spot. *Plant Pathol.* **57**, 518–526.
- Scott, P.R. (1971) The effect of temperature on eyespot (*Cercospora herpotrichoides*) in wheat seedlings. *Ann. Appl. Biol.* **68**, 169–175.
- Shagimardanova, E.I., Gusev, O.A., Sychev, V.N., Levinskikh, M.A., Sharipova, M.R., Il'inskaya, O.N., Bingham, G. and Sugimoto, M. (2010) Expression of stress response genes in barley *Hordeum vulgare* in a spaceflight environment. *Mol. Biol.* **44**, 734–740.
- Stabentheiner, E., Minihofer, T. and Huss, H. (2009) Infection of barley by *Ramularia collo-cygni*: scanning electron microscopic investigations. *Mycopathologia*, **168**, 135–143.
- Sun, L., Zhang, H., Li, D., Huang, L., Hong, Y., Ding, X.S., Nelson, R.S., Zhou, X. and Song, F. (2013) Functions of rice NAC transcriptional factors, ONAC122 and ONAC131, in defense responses against *Magnaporthe grisea*. *Plant Mol. Biol.* **81**, 41–56.
- Taylor, J.M.G., Paterson, L.J. and Havis, N.D. (2010) A quantitative real-time PCR assay for the detection of *Ramularia collo-cygni* from barley (*Hordeum vulgare*). *Lett. Appl. Microbiol.* **50**, 493–499.
- Thirugnanasambandam, A., Wright, K.M., Havis, N., Whisson, S.C. and Newton, A.C. (2011) *Agrobacterium*-mediated transformation of the barley pathogen *Ramularia collo-cygni* with fluorescent marker tags and live tissue imaging of infection development. *Plant Pathol.* **60**, 929–937.
- Tufan, H.A., McGrann, G.R.D., Magusin, A., Morel, J.B., Miche, L. and Boyd, L.A. (2009) Wheat blast: histopathology and transcriptome reprogramming in response to adapted and nonadapted *Magnaporthe* isolates. *New Phytol.* **184**, 473–484.
- Uauy, C., Distelfeld, A., Fahima, T., Blechl, A. and Dubcovsky, J. (2006) A NAC gene regulating senescence improves grain protein, zinc, and iron content in wheat. *Science*, **314**, 1298–1301.
- Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A. and Speleman, F. (2002) Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biol.* **3**, research0034.0031–research0034.0011.
- Walters, D.R., Havis, N.D. and Oxley, S.J.P. (2008) *Ramularia collo-cygni*: the biology of an emerging pathogen of barley. *FEMS Microbiol. Lett.* **279**, 1–7.
- Walters, D.R., Avrova, A., Bingham, I.J., Burnett, F.J., Fountaine, J., Havis, N.D., Hoad, S.P., Hughes, G., Looseley, M., Oxley, S.J.P., Renwick, A., Topp, C.F.E. and Newton, A.C. (2012) Control of foliar diseases in barley: towards an integrated approach. *Eur. J. Plant Pathol.* **133**, 33–73.
- West, J.S., Townsend, J.A., Stevens, M. and Fitt, B.D.L. (2012) Comparative biology of different plant pathogens to estimate effects of climate change on crop diseases in Europe. *Eur. J. Plant Pathol.* **133**, 315–331.
- Wu, A., Allu, A.D., Garapati, P., Siddiqui, H., Dortay, H., Zanol, M.-I., Asensi-Fabado, M.A., Munne-Bosch, S., Antonio, C., Tohge, T., Fernie, A.R., Kaufmann, K., Xue, G.-P., Mueller-Roeber, B. and Balazadeh, S. (2012) *JUNGBRUNNEN1*, a reactive oxygen species-responsive NAC transcription factor, regulates longevity in *Arabidopsis*. *Plant Cell*, **24**, 482–506.
- Xia, N., Zhang, G., Sun, Y.F., Zhu, L., Xu, L.S., Chen, X.M., Liu, B., Yu, Y.T., Wang, X.J., Huang, L.L. and Kang, Z.S. (2010) *TaNAC8*, a novel NAC transcription factor gene in wheat, responds to stripe rust pathogen infection and abiotic stresses. *Physiol. Mol. Plant Pathol.* **74**, 394–402.
- You, J., Zong, W., Li, X., Ning, J., Hu, H., Li, X., Xiao, J. and Xiong, L. (2013) The SNAC1-targeted gene *OsSRO1c* modulates stomatal closure and oxidative stress tolerance by regulating hydrogen peroxide in rice. *J. Exp. Bot.* **64**, 569–583.
- Zhou, Y., Huang, W., Liu, L., Chen, T., Zhou, F. and Lin, Y. (2013) Identification and functional characterization of a rice NAC gene involved in the regulation of leaf senescence. *BMC Plant Biol.* **13**, 132.

## SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site:

**Fig. S1** Quantitative reverse transcription-polymerase chain reaction (qRT-PCR) confirmation of constitutive increase in *HvSNAC1* transcript levels in transgenic barley over-expression (OE) lines. Error bars indicate  $\pm 1SE$ . \*\*\* $P < 0.001$  and \*\* $P < 0.01$  for comparison of means of OE lines with Golden Promise (GP).

**Table S1** Quantitative reverse transcription-polymerase chain reaction (qRT-PCR) primers used in this study.